

Live birth from TESA-ICSI in a modified natural cycle – A case report

Swati Kumari, Gunjan Kacker

Vatsalya IVF Centre, Kathmandu, Nepal

Abstract

Low ovarian reserve affects 10% of all women seeking fertility treatment. Management of patients with diminished reserve is challenging to fertility experts as they respond poorly to ovarian stimulation, and often such patients resort to oocyte donation. The number of births from natural cycle oocytes with surgically extracted sperm is low, but the case presented here describes a successful case in which a live pregnancy was achieved. In the treatment of a 36-year-old female with bilateral antral follicular count of one to two, two M II oocytes were obtained after modified natural cycle. The husband's semen analysis showed severe infection with occasional dead sperms. The patient did not respond to antibiotics and due to unavailability of sperm on the day of oocyte extraction, the oocytes were vitrified. Sperms were subsequently extracted by TESA. The vitrified oocytes were warmed, and ICSI was performed on the same day. One eight-cell grade I embryo was formed on D3 which was transferred in a programmed hormone replacement cycle. Pregnancy was obtained and a healthy infant was delivered. Hence, in women with low ovarian reserve, the use of modified natural cycle may provide a good alternative to achieving pregnancy prior to considering oocyte donation.

Keywords: IVF, low ovarian reserve, natural cycle, TESA-ICSI

Address for correspondence: Dr. Swati Kumari, MD Gynae/Obs, Vatsalya Natural IVF, Narayanchaur, Naxal, Kathmandu 44600, Nepal.
E-mail: drswatikumari88@gmail.com


INTRODUCTION

Low ovarian reserve is the reduced ability of the ovaries to produce oocytes and affects approximately 10% of women seeking fertility treatment.^[1] This condition is used to characterize women at risk for poor performance with assisted reproductive technologies.^[2-4]

The quantity and quality of ovarian follicles decrease with age.^[5] Different trials show that a woman has maximum fertility potential in the early 20s.^[5] The progressive decline of fertility increases in the late 30s, ending on average age of 50 to 51.^[6] The reduction of ovarian

function with ageing has been widely defined in terms of progressive reduction of ovarian follicles and diminished capability to generate competent oocytes.^[7,8] A premature reduction of ovarian reserve can also occur in young age. This condition can be a consequence of chemotherapy, radiation or surgery.^[9] However, in 90% of the cases, it is found to be idiopathic.^[9]

Several studies show that antral follicle count (AFC) and ovarian volume are very effective in estimating the response to ovarian stimulation.^[10] Ovarian antral follicles are evaluated by transvaginal ultrasound at the beginning of the follicular phase of menstrual cycle, and

Access this article online	
Quick Response Code: 	Website: www.fertilityscienceresearch.org
	DOI: 10.4103/fsr.fsr_26_18

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

How to cite this article: Kumari S, Kacker G. Live birth from TESA-ICSI in a modified natural cycle – A case report. *Fertil Sci Res* 2018;5:33-6.

the total number of 2 to 10 mm follicles in both the ovaries represents the AFC.^[11] AFC with diameter larger than 2 mm are described as 'recruitable', and they are highly sensitive and responsive to gonadotropins.^[12]

Management of women with compromised ovarian reserve is challenging for fertility experts and generally the only option to conceive is represented by assisted reproduction technologies (ART).

Women with reduced ovarian reserve yield less oocytes have less embryos for transfer, and their chances of pregnancy are obviously lower.^[13] In accordance with European Society of Human Reproduction and Embryology (ESHRE) consensus,^[14] patients with reduced ovarian reserve should be defined as 'expected poor responders' to ovarian stimulation during ART cycles. In women with a greatly reduced ovarian reserve, the strategy that provides the greatest chance of pregnancy is represented by egg donation.^[15] Natural cycles too can be tried as it allows a natural oocyte selection and an improved embryo quality.^[15]

Testicular sperm aspiration or TESA is the most efficient method for retrieving sperm.^[16] It requires special expertise and offers a less traumatic method for sperm retrieval.^[16] The first attempt at sperm retrieval is the best chance the patient has and gives the highest chance of success with the least morbidity.^[16] Fertilization with surgically obtained testicular sperms by intracytoplasmic sperm injection (ICSI) has been successfully used to achieve pregnancies in men with azoospermia.^[17] In obstructive azoospermia, 96% recovery rates have been reported using TESA.^[17] In non-obstructive azoospermia, reported recovery rates are approximately 50%,^[18] although fertilization rates are significantly lower in these men.^[19]

Before the advent of ICSI, donor sperm was used for successful fertilization.^[20] TESA followed by ICSI circumvents sperm donation in most cases.^[20]

As of date, there have been only a few reported cases of TESA followed by ICSI on vitrified oocytes from a modified natural cycle in a patient with low ovarian reserve. Hence, we present a very rare case in which successful pregnancy was achieved and a healthy live infant was born at term.

CASE REPORT

A 34-year-old, regularly menstruating female, married for 3.5 years, presented with a complain of primary infertility.

On transvaginal sonography, the antral follicular count was found to be one to two on both the sides, which was correlated with anti-mullerian hormone (AMH) of 1.38 ng/ml. Hormonal profile on D2 was thyroid stimulating hormone (TSH) 1.73 μ IU/ml, prolactin 9.62 ng/ml, follicle stimulation hormone (FSH) 6.8 mIU/ml, luteinizing hormone (LH) 5.23 mIU/ml and oestradiol 41.36 pg/ml.

The semen analysis of the male partner demonstrated 0.2 million non-motile dead sperms with plenty of leucocytes. The hormonal profile of the male partner was within normal range as follows: FSH 4.45 mIU/ml, testosterone 142 ng/dl. Antibiotics were prescribed in the hope of resolving the infection and improving the sperm count by the time of oocyte retrieval.

Meanwhile, the patient was stimulated with modified natural cycle and transvaginal follicular monitoring.

She was started on 100-mg clomiphene citrate on the second day of the cycle and on the sixth day of stimulation the dose was reduced to 50 mg when two to three oocytes were observed bilaterally. It was stopped on day 8. On the ninth day of stimulation, the follicular size was 15, 17 and 19 mm, respectively, and Inj. Cetrorelix 0.25 mg [Gonadotropin-releasing hormone (GnRH) antagonist] was started along with human menopausal gonadotropin (hMG) 150 IU. The patient was triggered with Inj. Deca 0.3 mg (GnRH agonist) when the three leading follicles attained the size of >18 mm. Ovum pick up (OPU) was performed 35 h later. Two M II oocytes were obtained. However, the male partner was not able to produce a semen sample on the day of oocyte retrieval due to high grade fever. Hence, ICSI could not be performed and the M II oocytes were vitrified 2 h after retrieval. The Cryotop method for oocyte vitrification was used. An improvement in semen parameter was expected after treatment with antibiotics, but a repeat semen sample was performed after a month demonstrated azoospermia. So, surgical retrieval of sperm was planned. In azoospermia, there will be no sperm in the epididymis. Hence, testicular sperm retrieval was planned at a later date.

A programmed hormone replacement cycle was started for the patient in the next cycle from day 3 of menses. She was started on oral E2 (tablet progynona, oestradiol valerate) 4 mg and the dose was increased to 6 mg on day 6. On the ninth day of endometrial preparation, the endometrial thickness was found to be 11 mm, and a triple-line pattern was observed on ultrasound. The patient was started on 800 mg of vaginal progesterone

suppositories in two divided doses daily for 3 days and TESA was performed. Few viable sperms were obtained and, ICSI was performed after warming the vitrified oocytes. One eight-cell grade I embryo was formed on day 3 and transferred using Labotect cannula, under abdominal ultrasound guidance.

Oral E2, vaginal progesterone, injectable progesterone, low-dose aspirin, multivitamins, folic acid and antacids were prescribed, and the woman was called after 14 days for urine pregnancy test. On the 14th day, the urine pregnancy test was found to be positive which was confirmed with serum beta human chorionic gonadotropin (β -hCG) of 526.0 mIU/ml. The hormonal supplementation was continued until 12 weeks of gestation. She had an uneventful antenatal period and delivered at term by normal vaginal delivery. She gave birth to a healthy 2.8-kg female baby.

DISCUSSION

The first report that investigated the natural cycle as a first strategy in women with low prognosis related to age and diminished ovarian reserve. It was performed by Papaleo *et al.* They found a similar pregnancy rate to that of conventional in vitro fertilization (IVF) in older women.^[21] Schimberni *et al.* evaluated 500 natural cycles followed by ICSI in poor responders. They observed that natural cycle IVF to be an efficacious protocol for poor responders, especially younger women.^[22]

However, the use of natural cycles presents some disadvantages mainly due to the frequent spontaneous LH surge, the resulting high cancellation rate (up to 30%), the difficulties in programming oocytes retrievals, the high incidence of failure to recover oocytes during oocyte pick-up (16.7%–71.4%) and low pregnancy rate per embryo transfer cycle (0%–23.5%).^[23]

Hence, modified natural cycle may be a more prudent approach to patients with low ovarian reserve. Modified natural cycle protocol means GnRH antagonist administration when a follicle attains 14mm on ultrasound, with the daily addition of FSH or hMG, to support additional follicular growth.^[24]

Yoo *et al.* compared IVF outcomes of mild ovarian stimulation with conventional ovarian stimulation in poor responders. They found a higher pregnancy rate in women who were recommended modified natural cycle stimulation protocols.^[25] In addition, modified natural

cycles IVF is a relatively easy procedure, especially for patients, that minimizes physical and emotional stress, the costs of treatment and laboratory tests.^[26] Natural cycle IVF allows a natural oocyte selection and an improved embryo quality.^[27] A case report by Akagbosu *et al.*^[28] and a paper by Aboulghar *et al.*^[29] suggest a negative impact of conventional IVF on oocyte quality. So, in our patient, our first approach was to try a modified natural cycle before considering other approaches, and we found that we were able to recruit three follicles and obtained two good-quality oocytes (M II).

The quality of the oocyte was confirmed by a good embryo formed even with TESA sperm. A study by Kalliopi *et al.* found that fertilization and cleavage rates, quality of embryos as well as blastocyst development rates were significantly reduced with TESA sperms.^[30] A possible explanation for this may be that testicular spermatozoa are less mature and subsequently less competent to fertilize than the ejaculated ones, as the final step of sperm maturation takes place in the epididymis.^[31] Embryo development can also be influenced by the quality of DNA in the sperm head. Testicular sperms have abnormal chromatin packing in the sperm head and hence lead to abnormal chromatin decondensation at fertilization.^[16,32] It is speculated that oocytes have the ability to correct small-scale DNA damage upon fertilization.^[32] The eggs obtained in our minimal stimulation natural cycle gave a good embryo and a live pregnancy even with a TESA sperm.

CONCLUSION

A good number of women with low ovarian reserve may conceive with their own eggs, if they are stimulated by modified natural cycle. Modified natural cycle yields good-quality eggs and embryos as compared to stimulated cycles in patients with low ovarian reserve and hence should be tried prior to considering egg donation. It is cost effective and has less psychological and financial burden for the patient.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

REFERENCES

1. Jirge PR. Poor ovarian reserve. *J Hum Reprod Sci* 2016;9:63.

2. Muasher SJ, Oehninger S, Simonetti S, Matta J, Ellis LM, Liu HC, *et al.* The value of basal and/or stimulated serum gonadotropin levels in prediction of stimulation response and in vitro fertilization outcome. *Fertil Steril* 1998;50:298-307.
3. Toner JP, Philput CB, Jones GS, Muasher SJ. Basal follicle-stimulating hormone level is a better predictor of in vitro fertilization performance than age. *Fertil Steril* 1991;55:784-91.
4. Esposito MA, Coutifaris C, Barnhart KT. A moderately elevated day 3 FSH concentration has limited predictive value, especially in younger women. *Hum Reprod* 2002;17:118-23.
5. Wood JW. Fecundity and natural fertility in humans. *Oxf Rev Reprod Biol* 1989;11:61-109.
6. te Velde ER, Pearson PL. The variability of female reproductive ageing. *Hum Reprod Update* 2002;8:141-54.
7. Sauer MV, Paulson RJ, Lobo RA. A preliminary report on oocyte donation extending reproductive potential to women over 40. *N Engl J Med* 1990;323:1157-60.
8. Ottolenghi C, Uda M, Hamatani T, Crisponi L, Garcia JE, Ko M, *et al.* Aging of oocyte, ovary, and human reproduction. *Ann N Y Acad Sci* 2004;1034:117-31.
9. Rasool S, Shah D. Fertility with early reduction of ovarian reserve: The last straw that breaks the Camel's back. *Fertil Res Pract* 2017;3:15.
10. Bancsi LF, Broekmans FJ, Eijkemans MJ, de Jong FH, Habbema JD, te Velde ER. Predictors of poor ovarian response in in vitro fertilization: A prospective study comparing basal markers of ovarian reserve. *Fertil Steril* 2002;77:328-36.
11. Broekmans FJ, de Ziegler D, Howles CM, Gougeon A, Trew G, Olivennes F. The antral follicle count: Practical recommendations for better standardization. *Fertil Steril* 2010;94:1044-51.
12. La Marca A, Spada E, Sighinolfi G, Argento C, Tirelli A, Giulini S, *et al.* Age-specific nomogram for the decline in antral follicle count throughout the reproductive period. *Fertil Steril* 2011;95:684-8.
13. Loutradis D, Vomvolaki E, Drakakis P. Poor responder protocols for in-vitro fertilization: Options and results. *Curr Opin Obstet Gynecol* 2008;20:374-8.
14. Ferraretti AP, La Marca A, Fauser BC, Tarlatzis B, Nargund G, Gianaroli L, *et al.* ESHRE consensus on the definition of 'poor response' to ovarian stimulation for in vitro fertilization: The Bologna criteria. *Hum Reprod* 2011;26:1616-24.
15. Maclaran K, Panay N. Premature ovarian failure. *J Fam Plann Reprod Health Care* 2011;37:35-42.
16. Shah R, Gupta C. Advances in sperm retrieval techniques in azoospermic men: A systematic review. *Arab J Urol* 2018;16:125-31.
17. Lindheim SR, Crumm K, Fisch H, Sauer MV. Testicular sperm aspiration (TESA) and its application in oocyte donation. *Arch Androl* 2001;46:211-5.
18. Levin HS. Testicular biopsy in the study of male infertility: Its current usefulness, histologic techniques, and prospects for the future. *Hum Pathol* 1979;10:569-84.
19. Craft I, Tsirigotis M, Bennett V, Taranissi M, Khalifa Y, Hogewind G, *et al.* Percutaneous epididymal sperm aspiration and intracytoplasmic sperm injection in the management of infertility due to obstructive azoospermia. *Fertil Steril* 1995;63:1038-42.
20. Sauer MV, Paulson RJ, Francis MM, Macaso TM, Lobo RA. Preimplantation adoption: Establishing pregnancy using donated oocytes and spermatozoa. *Hum Reprod* 1995;10:419-22.
21. Papaleo E, De Santis L, Fusi F, Doldi N, Brigante C, Marelli G, *et al.* Natural cycle as first approach in aged patients with elevated follicle-stimulating hormone undergoing intracytoplasmic sperm injection: A pilot study. *Gynecol Endocrinol* 2006;22:351-4.
22. Schimberni M, Morgia F, Colabianchi J, Giallonardo A, Piscitelli C, Giannini P, *et al.* Natural-cycle in vitro fertilization in poor responder patients: A survey of 500 consecutive cycles. *Fertil Steril* 2009;92:1297-301.
23. Ata B, Yakin K, Balaban B, Urman B. Embryo implantation rates in natural and stimulated assisted reproduction treatment cycles in poor responders. *Reprod Biomed Online* 2008;17:207-12.
24. Pelinck MJ, Vogel NE, Hoek A, Arts EG, Simons AH, Heineman MJ. Minimal stimulation IVF with late follicular phase administration of the GnRH antagonist cetrorelix and concomitant substitution with recombinant FSH: A pilot study. *Hum Reprod* 2005;20:642-8.
25. Yoo JH, Cha SH, Park CW, Kim JY, Yang KM, Song IO, *et al.* Comparison of mild ovarian stimulation with conventional ovarian stimulation in poor responders. *Clin Exp Reprod Med* 2011;38:159-63.
26. Janssens RMJ, Lambalk CB, Vermeiden JPW, Schats R, Schoemaker J. In-vitro fertilization in a spontaneous cycle: Easy, cheap and realistic. *Hum Reprod* 2000;15:314-8.
27. Munne S, Magli C, Adler A, Wright G, de Boer K, Mortimer D, *et al.* Treatment-related chromosome abnormalities in human embryos. *Hum Reprod* 1997;12:780-4.
28. Akagbosu F, Marcus S, Abusheikha N, Avery S, Brinsden P. Does ovarian hyperstimulation syndrome affect the quality of oocytes? *Hum Reprod* 1998;13:2583-4.
29. Aboulghar MA, Mansour RT, Serour GI, Ramzy AM, Amin YM. Oocyte quality in patients with severe ovarian hyperstimulation syndrome. *Fertil Steril* 1997;68:1017-21.
30. Loutradi K, Tarlatzis B, Goulis D, Zepiridis L, Pagou T, Chatziioannou E, *et al.* The effects of sperm quality on embryo development after intracytoplasmic sperm injection. *J Assist Reprod Gen* 2006;23:69-74.
31. Vernaev V, Tournaye H, Osmanagaoglu K, Verheyen G, Van Steirteghem A, Devroey P. Intracytoplasmic sperm injection with testicular spermatozoa is less successful in men with nonobstructive azoospermia. *Fertil Steril* 2003;79:529-33.
32. Sakkas D, Mariethoz E, Manicardi G, Bizzaro D, Bianchi PG, Bianchi U. Origin of DNA damage in ejaculated human spermatozoa. *Rev Reprod* 1999;4:31-7.