ScientificScholar[®] Knowledge is power Publisher of Scientific Journals https://fertilityscienceresearch.org



Fertility Science and Research



Review Article Fertility Problems Due to Chrono-Disruption: A Mini Review

Megha Das¹, Sanjeev Kumar Yadav¹, Nitesh Kumar Mishra¹, Chandana Haldar¹

¹Department of Zoology, Banaras Hindu University, Varanasi, India.



***Corresponding author:** Prof. Chandana Haldar, MSc, PhD, Department of Zoology, Banaras Hindu University, Varanasi, India

chaldarbhu@gmail.com

Received: 14 May 2024 Accepted: 09 October 2024 Published: 28 November 2024

DOI 10.25259/FSR_11_2024

Quick Response Code:



ABSTRACT

Circadian rhythm coordinates many physiological and behavioural processes and is under the control of the endogenous suprachiasmatic nucleus (SCN) entrained by a light-dark cycle. The core clock genes, *Bmal1*, *Clock*, *Per*, and *Cry*, organised are organised in a transcriptional–translational feedback loop (TTFL) of the 24-h cycle. Recently, these clocks have been reported in the female reproductive organs, i.e., ovary, uterus, etc. The presence of clock genes in female reproductive tissues has generated the interest of reproductive biologists towards chrono-regulation, as the clock genes may play a central role in coordinating the circadian rhythm of the reproductive process, i.e., form ovulation, fertilisation and implantation to lactation. Studies have demonstrated a critical connection between disturbance of the master circadian clock (SCN) of the hypothalamus and infertility.

Artificial lighting is a worldwide increasing problem in the physiology of many vertebrates controlled by circadian/seasonal entrainments. The effect of artificial light on circadian dysregulation in the night-shift working females resulted in severe reproductive dysfunction (low pregnancy outcomes, childbirth weight, multiple miscarriages, etc.). Inappropriate exposure to artificial light/chrono disruption can lead to several disturbances to the biological/physiological rhythms of metabolomics and hormones.

In the present mini review, we proposed that the duration of light affects the circadian coordination of central (SCN) and peripheral clock (ovarian and uterine clock) oscillations regulating uterine tissue homeostasis and pregnancy success in golden hamsters. Two major aspects of light exposure, i.e., (1) continuous and (2) long-time artificial light at night (ALAN), influence circadian oscillations of clock genes on SCN, ovary, and uterus as well as the molecular mechanism behind the altered clock entrainments that finally induce the female reproductive impairments via alteration in hypothalamic-pituitary-gonadal (HPG) axis.Keywords: Smoking, Semen Quality, Sperm Parameters, Male Infertility, North India, Fertility Treatment, Smoking Cessation.

Keywords: Chronodisruption, Artificial light, Clock genes, Female infertility, Circadian

INTRODUCTION

Circadian Clock - SCN

The circadian clock, or the suprachiasmatic nucleus (SCN), is a molecular timekeeping machine located in the anterior hypothalamus at the dorsal side of the optic chiasma.^[1] SCN is the central circadian pacemaker consisting of a cluster of 20,000 excitatory neurones and astrocytes, whose activities fluctuate with the 24-h cycle. It participates in diverse photic (light) and non-photic (metabolic, temperature) stimuli for the regulation of physiological activities, which include the sleep/wake cycle, energy metabolism, hormonal regulation, immune functions, and cell proliferation.^[2] Retinal light information from the retinohypothalamic tract (RHT) reaches the SCN and communicates periodically with non-SCN secondary brain clocks and then with

This is an open-access article distributed under the terms of the Creative Commons Attribution-Non Commercial-Share Alike 4.0 License, which allows others to remix, transform, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms. ©2024 Published by Scientific Scholar on behalf of Fertility Science and Research

peripheral organ clocks (e.g., heart, liver, kidney, ovary, uterus, adrenal, etc.) to synchronise with external day and night cycles.^[2] This master clock, SCN, rhythmically coordinates organismal physiology and behaviour in response to daily environmental changes by synchronising non-SCN subordinate brain clocks and peripheral organ clocks by neural (neurotransmitters and neuropeptides) and humoral outputs (hormonal secretion, neural innervations, and autonomic nervous system).^[3]

CIRCADIAN CONTROL ON FEMALE FERTILITY

Circadian Rhythm and Ovulation

Ovulation is the interplay of various hormones that is crucial to making ovulation successful. The circadian system regulates the HPG axis, which in turn plays a prominent role in the management of steroid hormone synthesis, follicular development and ovulation till maturation.^[4,5] Successful pregnancy commences with the release of mature oocyte from the ovary and terminates with parturition. Rhythmical expression of kisspeptin (Kiss1) activates the gonadotropin-releasing hormone (GnRH) neurons.^[6] At the mature pre-ovulatory stage, the GnRH secretion becomes stimulatory and simultaneously excites the anterior pituitary to release LH, leading to the discharge of mature oocytes from the ovary. A wide array of studies regarding the circadian regulation of female reproduction is confined to the hormonal crosstalk between SCN and LH surge.^[7]

To understand the role of clock genes on the ovulation process, an assessment was conducted by the researchers, where the expression of aryl hydrocarbon receptor nuclear translocator-like protein (Arntl) or Bmal1 and Per2 was checked over 2 days during the oestrous cycle, which suggested that the rhythm of circadian clock genes of the ovary might be regulated significantly by LH secretion.^[8] Reports also suggested that in granulosa cells (GCs) several genes are clock-controlled such as LH receptor (Lhcgr), prostaglandin synthesis enzymes (Ptgs2), steroidogenic enzymes (e.g., Cyp11a1, aromatase, etc.), delta-aminolevulinate synthase 1 (Alas1), Ppary coactivator 1 alpha (Pparyc1α), interleukin 6 (IL6), and gap junction protein Connexin-43 (Cx43) that are important in follicular development and ovulation.^[9] It has also been stated that the clock genealtered estradiol signalling.^[10] Further, the promoter region of cyclooxygenase-2 (COX2), a rate-limiting enzyme for prostaglandin synthesis, has an E-box sequence which binds to the CLOCK: BMAL1 heterodimers and activates its rhythmic transcription during follicular maturation.^[7] Further, this rhythmic accumulation of COX2 enzyme regulates the rhythmic synthesis of prostaglandin E2 (PGE2) and prostaglandin F2a (PGF2a), resulting in increased levels of prostaglandin synthesis.^[7]

Circadian Rhythm and Fertilization

The fertile window initiates approximately between 3 and 6 days before LH and remains till the deposited sperms remain viable. After 12–48 h of the LH surge, ovulation occurs.^[11,12] It has been observed that for successful reproductive outcomes, mating should occur early, just after ovulation.^[13] As the level of estradiol increases by 2–3 days before the LH surge, the mucus layer of the reproductive tract becomes permeable due to an increase in hydration, thus allowing sperms to travel into the uterus.^[14]

Subsequently, the timing of LH surge and estradiol level creates an important *milieu* for sperm motility and fertilisation capability.^[11] It was well established that clock genes are responsible for estradiol synthesis and LH surge; thus, it can be proposed that the fertilisation capacity is also indirectly regulated by circadian timing.^[11] Studies with hypophysectomised juvenile rats confirmed no such circadian rhythm pattern of *Arntl* and *Per2* due to the absence of LH surge. Further, in mice, a lack of both *Cry1* and *Cry2* was noted in the absence of LH surge.^[8,15–17] Therefore, hormonal coordination with clock gene oscillations is important for ovulation and fertilisation.

Circadian Rhythm and Implantation

Implantation is a process of adherence of the blastocyst to the receptive endometrium, which includes the series of changes in the uterine environment guided by ovarian E2 and P4 to establish mutual signalling between the blastocyst and the uterus.^[18,19] During the period of uterine receptivity, the clock genes and their proteins express rhythmically within the uterine epithelium, stroma, and myometrium.^[20-22] It has been reported that ovarian steroid hormones have the capability to change the expression of clock genes.^[23] Progesterone not only increases the expression of neuronal PAS domain protein 2 (Npas2), Clock, Cry1, and Per1 expression but also decreases Rev-erbß and retinoic acid receptor-related orphan receptor-a (RORg) mRNA expression.^[24] Additionally, over the peri-implantation period, the vascular endothelial growth factor (VEGF) mRNA (an E-box containing mRNA at its promoter) also expresses rhythmically in the uterus.^[20] A knockout study showed that implantation failure in Bmal1 knockout mice is due to a lack of P4 biosynthesis enzyme expression. In the luteinised GCs, Per1 and Per2 mutant mice showed 80% fertility.^[22, 25, 26] Studies confirmed that women with polymorphic Bmal1, Bmal2, Clock, and Npas2 express normal female reproductive characteristics.[27] The report also suggested that the Bmal1 gene (rs2278749 TT) contains SNP in association with an increased number of pregnancies as well as an increased number of miscarriages. Interestingly, they also revealed that Npas2 rs11673746 T carriers showed lower miscarriages.^[27] Hence, it can be concluded that implantation is also crucially governed by the circadian clock system.

Clock Rhythm and Embryogenesis

Reports suggest clock genes' rhythmic expression in unfertilised oocytes, pronuclear zygotes, blastocysts, and embryos.^[28] Further, CLOCK expression was noted at a high level throughout the pregnancy, starting from fertilisation. Studies have established that the incidence of all core clock genes mRNA expression occurs at various stages of oocytes and pre-implantation development of mouse and rabbit embryos.^[29] The uterus, placenta, and membrane of day 16 (D16) embryos were reported to have a stout rhythm of clock genes.^[24] Per3 has also been reported in association with corticogenesis, in which PER3 regulates excitatory neurone migration and synaptic network formation.^[30] An in vitro study of human embryonic stem cells (ESCs) showed that all clock genes are expressed in embryonic stem cells but not in a rhythmic way; thus, clock genes possess an important role in embryogenesis.^[31]

Circadian Rhythm and Gestation

Studies have revealed that the circadian timing of birth depends not only on the master clock (SCN) but also on the peripheral clocks. These clock genes participate in several reproductive peripheral tissues at the time of gestation, revealing that circadian rhythmicity associated with clock genes is important for parturition.^[32] The SCN lesion in rats showed a single peak in parturition frequency while in normal conditions, rats delivered during a 36-h time window with two peaks in parturition frequency 24 h apart.^[32] *Clock-/-* study revealed that pregnant mice lacking the *Clock* genes not only showed oa high incidence of foetal resorption but also had prolonged and unproductive labour.^[33,34] Some case studies have revealed that night time-delivered mice lack *Bmal1* in the myometrium.^[35] These reports suggested that clock gene oscillation is essential for healthy labour.

Circadian Rhythm and Female Hormonal Control

LH and FSH

The master clock communicates with several peripheral clocks via various but poorly defined connections of neural and humoral centres. After receiving the environmental signal, the core clock genes of SCN (*Bmal, Clock, Per*, and *Cry*) send the timing information via efferent neurones to the other part of the brain, mainly paraventricular nucleus (PVN), medial pre-optic area, dorsomedial nucleolus of hypothalamus, and the pineal gland (MEL), and regulate the secretion of hypophyseal hormones and melatonin.^[36] SCN is reported to synchronise the GnRH neurones in the medial preoptic area (MPOA).^[37]

Estrogen (E₂) & progesterone (P₄)

 E_2 and P_4 of the HPG axis are mainly responsible for all the molecular events related to pregnancy, such as ovulation, fertilisation, uterine receptivity, implantation, decidualisation, gestation, and parturition.^[38] Steroids influence the phase, amplitude, and period of circadian clock gene rhythms in SCN.^[38] The estrogen receptors (ER α , ER β) possess direct links with core clock genes.^[39] The promoter of ER β covers an evolutionary conserved E-box, which is the binding site of CLOCK/BMAL1 heterodimer in an arhythmic manner and regulates the rhythmic expression of the receptor.^[40,41] A complex loop relationship between estradiol signalling and clock proteins exists where estradiol may impact the core clock machinery.

HPA Axis

The HPA axis manifests all the maternal-foetal physiological adaptations for a successful pregnancy. On the contrary, the high foetal demand are for energy and the maternal physiological stress management are fulfilled by the HPA axis adaptation, which in turn promotes the release of the energy store as well as regulates maternal-foetal immune parameters.^[42,43] Moreover, reports suggested that in the SCN, clock genes express rhythmic controls on the circadian variations of both the HPA axis and the respective gonadal and adrenal peripheral clocks.^[44] Literature suggests that balance in sleep and wakefulness preserves both the body clock and a good melatonin level that keeps the body away from chrono-disruption-related diseases. The work at night/ shift work increases the time/duration of light exposure at night, consequently suppressing melatonin production, hence, potentially elevating cancer risk and the substantial downstream molecular mechanisms of the circadian clock genes with melatonin.

The destruction of the master clock through any environmental or lifestyle shifts may abolish the peripheral clock synchronisation and disrupt maternal physiological adjustments during pregnancy. We provided the fundamental explanation of the much-required anthropological perspective of the hazardous effects of light, and it may expand our understanding of the duration and spectral composition of light is essential for coordination between central and peripheral clock genes to regulate female reproductive health, including pregnancy.

Concept of chronodisruption

In 2003, the term "chronodisruption" was first invented_by T. C. Erren, R. J. Reiter, and C. Piekarski of the University of Cologne.^[45] Previously, desynchronisation 24-, there was a concept of "chronodisturbance" (or absence of adverse consequences for health), "circadian disrupt", or "disruption

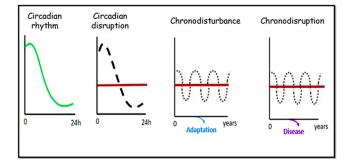


Figure 1: Concept of circadian disruption, chronodisturbance and chronodisruption (Modified from https://doi.org/10.3390/toxins12030151).

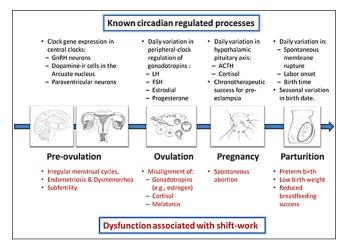


Figure 2: Reproductive impairment associated with shift work. LH: Luteinizing hormone, FSH: Follicle Stimulating hormone, ACTH: Adrenocorticotropic hormone. (Modified from https://doi. org/10.3389/fendo.2013.00092).

of circadian rhythm," suggesting the desynchronisation of 24-h rhythms on adverse health. Chronodisruption was further classified as "exogenous and endogenous exposures which can disrupt the timing and order of physiologic functions."^[45] The nighttime use of artificial light and backlit screens during the night is a very prominent example of a chronodisrupter.^[46] The International Agency for Research on Cancer 2007 classified shift work as a chronodisrupter and also as a probable human carcinogen, and since then, "chronodisruption" has drawn the attention of the scientific world [Figure 1].^[46]

Light at Night Induced Chronodisruption and Female Fertility

In humans, it has been reported that circadian disruption can possess deleterious effects on female reproduction that are linked to female hormonal systems, resulting in reproductive dysfunction-related subfertility.^[47] Studies have suggested the presence of multiple targets of the molecular

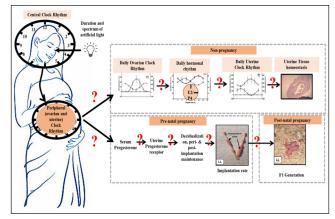


Figure 3: Showing the effect of duration of light in desynchronization of central clock and clocks in reproductive organs (Modified from https://doi.org/10.1007/s43630-022-00210-6).

loop of the circadian clock that is associated with important physiological functions of the hypothalamus-pituitarygonadal axis, resulting in disturbed female reproduction.^[48] Previously, survey studies on hospital nurses showed that night shift or rotating shift nurses experience painful and irregular menstruation with changes in their cycle length, duration of bleeding period, menstrual flow, and dysmenorrhea.^[49,50] Same group of researchers showed that hospital nurses in shift work had irregular menstrual cycles than permanent day shift working nurses.^[49,50] The expansion of the menstrual cycle is connected to the length of the follicular phase. These results suggest that rotating shift work may induce a delay in ovulation.^[50] Further, studies on knockout or transgenic animal models have shown that clock gene synchronisation is critical for reproductive success in female rodents. Circadian dysregulation in clock genes is also known to disrupt pregnancy, such as time of gestation, circadian timing of birth, etc. [Figure 2].^[48,51]

The biological rhythms of humans are reported to be affected by constant light.^[52] Several studies have revealed that the biological crosstalk between the mother and foetus is affected by constant light. Therefore, disturbances in the ratio of light and dark not only disturb the pattern of sleep in such as the mother but also affect the early stages of various development, such as visual development of the foetus, postnatal weight gain, etc.^[53–57]

Our studies with mice suggested that in reproductive tissue, continuous light (LL) damages the circadian coordination between central and clock genes, leading to disturbed uterine physiology and, thus, altered pregnancy. Studies on pregnant mice revealed lowered progesterone under ;the LL condition; mice under the LL condition downregulated expression of progesterone receptor (PR) and PR-dependent uterine Homeobox A-10 (Hoxa10) proteins. It lowered pregnancy outcomes in LL mice. Hence, we may propose that

in females, the duration of light exposure at the workplace/ home is important to minimise pregnancy-related problems [Figure 3].^[58]

LL is always negatively affecting the reproductive health of females. LL exposure increases the endometrium thickness and reduces the myometrium, as the condition of uterine adenomyosis. LL alters the expressions of clock genes in SCN, ovary, and uterus along with serum estradiol rhythm gets disturbed as well in non-pregnant females *Egf* gets upregulated, and Aanat, Cx26, and Cx43 mRNA levels get downregulated in the uterus. LL exposure desynchronises the central and peripheral reproductive clock and thus uterine physiology via the Akt/FoxO1 pathway, as reported in Golden Hamsters.^[59] Hence, LL could be a risk factor for female fertility. Thus, the duration of the light is important in considering the physiological consequences of female reproductive abnormalities.

CONCLUSION

Our review may provide a fundamental explanation of the much-required anthropological perspective of the hazardous effects of light. It may expand our understanding of the duration and spectral composition of light that are essential in coordination between central and peripheral clock genes in the regulation of female reproductive health and pregnancy success that minimises the risk factor for females at the workplace/home. ALAN is unavoidable in the current scenario. Not only streetlights, but the use of electronic devices is increasing day by day which also emits bluewavelength-rich light. To find out the future possibilities to restore a healthy pregnancy in the present urbanised world, it is necessary to understand the cause and the mechanism involved in the prevention of normal physiological processes of pregnancy. However, the information is still not enough regarding the spectrum and intensity of light on the clock regulation of pregnancy impairments like unsuccessful implantation/spontaneous abortions. More studies are required to delineate the molecular mechanism of clock genes and the effect of light/light spectrum on chronodisruptive female fertility problems.

Acknowledgement

The authors would like to acknowledge the ICMR ad-hoc project (P-14/267) and Institute of Eminence (IoE) research grant, BHU, to Dr. Sanjeev K. Yadav.

Author contribution

Conception and design: MD, SKY, Collection and assembly of data: MD, NKM, Writing, review and editing: MD, SKY, CH, Final approval of manuscript: All authors.

Ethical approval

Institutional Review Board approval is not required.

Declaration of patients consent

Patient's consent not required as there are no patients in this study.

Financial support and sponsorship

The authors are thankful to the Indian Council of Medical Research and Institute of Eminence, BHU, for financial support to Dr. Sanjeev K. Yadav.

Conflicts of interest

There are no conflicts of interest.

Use of artificial intelligence (AI)-assisted technology for manuscript preparation:

The authors confirm that there was no use of Artificial Intelligence (AI)-Assisted Technology for assisting in the writing or editing of the manuscript and no images were manipulated using AI.

REFEENCES

- 1. Ma MA, Morrison EH. Neuroanatomy, Nucleus Suprachiasmatic. In: StatPearls. StatPearls Publishing; 2023.
- 2. Koronowski KB, Sassone-Corsi P. Communicating Clocks Shape Circadian Homeostasis. Science. 2021;371(6530):eabd0951.
- Buijs RM, Guzmán Ruiz MA, Méndez Hernández R, Rodríguez Cortés B. The Suprachiasmatic Nucleus: A Responsive Clock Regulating Homeostasis by Daily Changing the Setpoints of Physiological Parameters. Auton Neurosci. 2019;218:43–50.
- Gräs S, Georg B, Jørgensen HL, Fahrenkrug J. Expression of the Clock Genes Per1 and Bmal1 During Follicle Development in the Rat Ovary. Effects of Gonadotropin Stimulation and Hypophysectomy. Cell Tissue Res. 2012;350(3):539–48.
- Yoshikawa T, Sellix M, Pezuk P, Menaker M. Timing of the Ovarian Circadian Clock is Regulated by Gonadotropins. Endocrinology. 2009;150(9):4338–47.
- Zeydabadi Nejad S, Ramezani Tehrani F, Zadeh-Vakili A. The Role of Kisspeptin in Female Reproduction. Int J Endocrinol Metab. 2017;15(3):e44337.
- Sellix MT, Menaker M. Circadian Clocks in the Ovary. Trends Endocrinol Metab. 2010;21(10):628–36.
- Karman BN, Tischkau SA. Circadian Clock Gene Expression in the Ovary: Effects of Luteinizing Hormone. Biol Reprod. 2006;75(4):624–32.
- Bozek K, Relógio A, Kielbasa SM, Heine M, Dame C, Kramer A, et al. Regulation of Clock-Controlled Genes in Mammals. PLoS One. 2009;4(3):e4882.
- Urlep Z, Rozman D. The Interplay Between Circadian System, Cholesterol Synthesis, and Steroidogenesis Affects Various Aspects of Female Reproduction. Front Endocrinol (Lausanne). 2013;4:111.

- Baird DD, McConnaughey DR, Weinberg CR, Musey PI, Collins DC, Kesner JS, et al. Application of a Method for Estimating Day of Ovulation Using Urinary Estrogen and Progesterone Metabolites. Epidemiology. 1995;6(5):547–50.
- Luciano AA, Peluso J, Koch EI, Maier D, Kuslis S, Davison E. Temporal Relationship and Reliability of the Clinical, Hormonal, and Ultrasonographic Indices of Ovulation in Infertile Women. Obstet Gynecol. 1990;75(3 Pt 1):412–6.
- Nakao S, Ito K, Sugahara C, Watanabe H, Kondoh G, Nakagata N, et al. Synchronization of the Ovulation and Copulation Timings Increased the Number of in Vivo Fertilized Oocytes in Superovulated Female Mice. PLoS One. 2023;18(2):e0281330.
- Katz DF, Slade DA, Nakajima ST. Analysis of Pre-Ovulatory Changes in Cervical Mucus Hydration and Sperm Penetrability. Adv Contracept. 1997;13(2–3):143–51.
- 15. Shearman LP, Sriram S, Weaver DR, Maywood ES, Chaves I, Zheng B, et al. Interacting Molecular Loops in the Mammalian Circadian Clock. Science. 2000;288(5468):1013–9.
- Okamura H, Miyake S, Sumi Y, Yamaguchi S, Yasui A, Muijtjens M, et al. Photic Induction of mPer1 and mPer2 in Cry-Deficient Mice Lacking a Biological Clock. Science. 1999;286(5449):2531–4.
- 17. Vitaterna MH, Selby CP, Todo T, Niwa H, Thompson C, Fruechte EM, et al. Differential Regulation of Mammalian Period Genes and Circadian Rhythmicity by Cryptochromes 1 and 2. Proc Natl Acad Sci USA. 1999;96(21):12114–9.
- Zhang S, Lin H, Kong S, Wang S, Wang H, Wang H, et al. Physiological and molecular determinants of embryo implantation. Mol Aspects Med. 2013;34(5):939–80.
- Dey SK, Lim H, Das SK, Reese J, Paria BC, Daikoku T, et al. Molecular Cues to Implantation. Endocr Rev. 2004;25(3):341–73.
- 20. Uchikawa M, Kawamura M, Yamauchi N, Hattori MA. Down-Regulation of Circadian Clock Gene Period 2 in Uterine Endometrial Stromal Cells of Pregnant Rats During Decidualization. Chronobiol Int. 2011;28(1):1–9.
- 21. Nakamura TJ, Sellix MT, Kudo T, Nakao N, Yoshimura T, Ebihara S, et al. Influence of the Estrous Cycle on Clock Gene Expression in Reproductive Tissues: Effects of Fluctuating Ovarian Steroid Hormone Levels. Steroids. 2010;75(3):203–12.
- Ratajczak CK, Herzog ED, Muglia LJ. Clock Gene Expression in Gravid Uterus and Extraembryonic Tissues During Late Gestation in the Mouse. Reprod Fertil Dev. 2010;22(5):743–50.
- 23. He PJ, Hirata M, Yamauchi N, Hattori MA. Up-Regulation of Per1 Expression by Estradiol and Progesterone in the Rat Uterus. J Endocrinol. 2007;194(3):511–9.
- 24. Nakamura K, Inoue I, Takahashi S, Komoda T, Katayama S. Cryptochrome and Period Proteins are Regulated by the CLOCK/BMAL1 Gene: Crosstalk Between the PPARs/ RXRalpha-Regulated and CLOCK/BMAL1-Regulated Systems. PPAR Res. 2008;2008:348610.
- Pilorz V, Steinlechner S. Low Reproductive Success in Per1 and Per2 Mutant Mouse Females Due to Accelerated Ageing? Reproduction. 2008;135(4):559–68.
- Zheng B, Albrecht U, Kaasik K, Sage M, Lu W, Vaishnav S, et al. Nonredundant Roles of the mPer1 and the mPer1 and mPer2 Genes in the Mammalian Circadian Clock. Cell. 2001;105(5):683–94.

- 27. Kovanen L, Saarikoski ST, Aromaa A, Lonnqvist J, Partonen T. ARNTL (BMAL1) and NPAS2 Gene Variants Contribute to Fertility and Seasonality. PLoS One. 2010;5:10007.
- Johnson MH, Lim A, Fernando D, Day ML. Circadian Clockwork Genes are Expressed in the Reproductive Tract and Conceptus of the Early Pregnant Mouse. Reprod Biomed Online. 2002;4(2):140–5.
- 29. Amano T, Tokunaga K, Kakegawa R, Yanagisawa A, Takemoto A, Tatemizo A, et al. Expression Analysis of Circadian Genes in Oocytes and Preimplantation Embryos of Cattle and Rabbits. Anim Reprod Sci. 2010;121(3-4):225-35.
- Noda M, Iwamoto I, Tabata H, Yamagata T, Ito H, Nagata KI. Role of Per3, A Circadian Clock Gene, in Embryonic Development of Mouse Cerebral Cortex. Sci Rep. 2019;9(1):5874.
- Dierickx P, Vermunt MW, Muraro MJ, Creyghton MP, Doevendans PA, Van Oudenaarden A, et al. Circadian Networks in Human Embryonic Stem Cell-Derived Cardiomyocytes. EMBO Rep. 2017;18(7):1199–212.
- Reppert SM, Henshaw D, Schwartz WJ, Weaver DR. The Circadian-Gated Timing of Birth in Rats: Disruption by Maternal SCN Lesions or by Removal of the Fetal Brain. Brain Res. 1987;403(2):398–402.
- Dolatshad H, Campbell EA, O'Hara L, Maywood ES, Hastings MH, Johnson MH. Developmental and Reproductive Performance in Circadian Mutant Mice. Hum Reprod. 2006;21(1):68–79.
- Miller BH, Olson SL, Turek FW, Levine JE, Horton TH, Takahashi JS. Circadian Clock Mutation Disrupts Estrous Cyclicity and Maintenance of Pregnancy. Curr Biol. 2004;14(15):1367–73.
- Ratajczak CK, Asada M, Allen GC, McMahon DG, Muglia LM, Smith D, et al. Generation of Myometrium-Specific Bmal1 Knockout Mice for Parturition Analysis. Reprod Fertil Dev. 2012;24(5):759–67.
- Takahashi JS, Hong HK, Ko CH, McDearmon EL. The Genetics of Mammalian Circadian Order and Disorder: Implications for Physiology and Disease. Nat Rev Genet. 2008;9(10):764–75.
- Colledge WH. Kisspeptins and GnRH Neuronal Signalling. Trends Endocrinol Metab. 2009;20(3):115–21.
- Angelousi A, Kassi E, Nasiri-Ansari N, Weickert MO, Randeva H, Kaltsas G. Clock Genes Alterations and Endocrine Disorders. Eur J Clin Investig. 2018;48(6):e12927.
- Lee Y, Chun SK, Kim K. SUMOylation Controls CLOCK-BMAL1-Mediated Clock Resetting Via CBP Recruitment in Nuclear Transcriptional Foci. Biochim Biophys Acta. 2015;1853(10 Pt A):2697–708.
- Koike N, Yoo SH, Huang HC, Kumar V, Lee C, Kim TK, et al. Transcriptional Architecture and Chromatin Landscape of the Core Circadian Clock in Mammals. Science. 2012;338(6105):349–54.
- 41. Ripperger JA, Schibler U. Rhythmic CLOCK-BMAL1 Binding to Multiple E-box Motifs Drives Circadian Dbp Transcription and Chromatin Transitions. Nat Genet. 2006;38(3):369–74.
- 42. Wharfe MD, Mark PJ, Wyrwoll CS, Smith JT, Yap C, Clarke MW, et al. Pregnancy-Induced Adaptations of the Central Circadian Clock and Maternal Glucocorticoids. J Endocrinol. 2016;228(3):135–47.
- 43. Ruffaner-Hanson C, Noor S, Sun MS, Solomon E, Marquez LE, Rodriguez D. The Maternal-Placental-Fetal Interface: Adaptations of the HPA Axis and Immune Mediators

Following Maternal Stress and Prenatal Alcohol Exposure. Exp Neurol. 2022;355:114121.

- Nader N, Chrousos GP, Kino T. Interactions of the Circadian CLOCK system and the HPA axis. Trends Endocrinol Metab. 2010;21(5):277–86.
- Erren TC, Reiter RJ, Piekarski C. Light, Timing of Biological Rhythms, and Chronodisruption in Man. Naturwissenschaften. 2003;90(11):485–94.
- Carriazo S, Ramos AM, Sanz AB, Sanchez-Niño MD, Kanbay M, Ortiz A. Chronodisruption: A Poorly Recognized Feature of CKD. Toxins (Basel). 2020 Feb 28;12(3):151.
- 47. Sen A, Sellix MT. The Circadian Timing System and Environmental Circadian Disruption: From Follicles to Fertility. Endocrinology. 2016;157(9):3366–73.
- Hsu CN, Tain YL. Light and Circadian Signaling Pathway in Pregnancy: Programming of Adult Health and Disease. Int J Mol Sci. 2020;21(6):2232.
- 49. Feskanich D, Hankinson SE, Schernhammer ES. Nightshift Work and Fracture Risk: The Nurses' Health Study. Osteoporos Int. 2009;20(4):537–42.
- Labyak S, Lava S, Turek F, Zee P. Effects of Shiftwork on Sleep and Menstrual Function in Nurses. Health Care Women Int. 2002;23(6–7):703–14.
- Touitou Y, Reinberg A, Touitou D. Association Between Light at Night, Melatonin Secretion, Sleep Seprivation, and The Internal Clock: Health Impacts and Mechanisms of Circadian Disruption. Life Sci. 2017;173:94–106.
- Brandon DH, Holditch-Davis DD, Belyea M. Preterm Infants Born at Less Than 31 Weeks' Gestation Have Improved Growth in Cycled Light Compared with Continuous Near Darkness. J Pediatr. 2002;140(2):192–9.

- Kennedy KA, Fielder AR, Hardy RJ, Tung B, Gordon DC, Reynolds JD, et al. Reduced Lighting Does Not Improve Medical Outcomes in Very Low Birth Weight Infants. J Pediatr. 2001;139(4):527–31.
- 54. Fielder AR, Moseley MJ. Environmental Light and the Preterm Infant. Semin Perinatol. 2000;24(4):291–8.
- 55. Mirmiran M, Ariagno RL. Influence of light in the NICU on the Development of Circadian Rhythms in Preterm Infants. Semin Perinatol. 2000;24(4):247–57.
- Miller CL, White R, Whitman TL, O'Callaghan MF, Maxwell SE. The Effects of Cycled Versus Noncycled Lighting on Growth and Development in Preterm Infants. Infant Behav Dev. 1995;18(1):87–95.
- Mann NP, Haddow R, Stokes L, Goodley S, Rutter N. Effect of Night and Day on Preterm Infants in a Newborn Nursery: Randomised Trial. Br Med J (Clin Res Ed). 1986;293(6557):1265–7.
- Das M, Minocha T, Kumar D, Yadav SK, Haldar C. Continuous Artificial Light Potentially Disrupts Central and Peripheral Reproductive Clocks Leading to Altered Uterine Physiology and Reduced Pregnancy Success in Albino Mice. Photochem Photobiol Sci 21, 1217–1232 (2022).
- Gamble KL, Resuehr D, Johnson CH. Shift Work and Circadian Dysregulation of Reproduction. Front Endocrinol (Lausanne). 2013;4:92.

How to cite this article: Das M, Yadav SK, Mishra NK, Haldar C. Fertility Problems Due to Chronodisruption: A Mini Review. Fertil Sci Res. 2024;11:13. doi: 10.25259/FSR_11_2024